

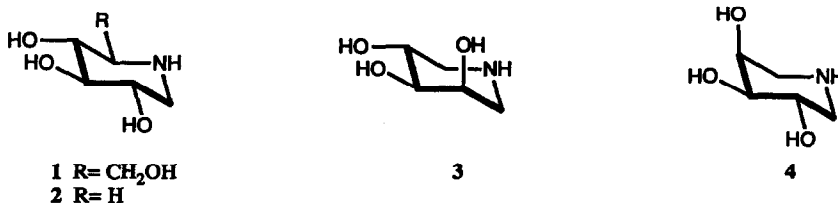
## A NEW FAMILY OF FIVE-CARBON IMINOALDITOLS WHICH ARE POTENT GLYCOSIDASE INHIBITORS

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**Abstract** – The synthesis of 1,5-dideoxy-1,5-imino-D-xylitol **2** (and the enantiomeric iminoalditols **3** and **4**) is described. Alditol **2** compares favorably with 1-deoxynojirimycin **1** as a potent inhibitor of  $\beta$ -glucosidase.

Deoxynojirimycin **1** effectively inhibits the enzymatic hydrolysis of  $\beta$ -D-glucosides.<sup>2</sup> The potential chemotherapeutic importance of such inhibitors of glucose metabolism<sup>3-7</sup> has prompted considerable synthetic interest in these alkaloids.<sup>8-10</sup> We reasoned that improved inhibitors might be rationally designed by examining how each hydroxyl substituent in the glycone (e.g. D-glucose) contributed to the free energy of binding with the enzyme. Dale *et al.* have systematically studied the inhibition of sweet almond  $\beta$ -glucosidase by a wide variety of normal and deoxysugars.<sup>11</sup> While the stereochemical configurations of individual ring hydroxyls were important, removing the C6-hydroxymethyl substituent altogether had remarkably little effect on enzyme-substrate interactions. This surprising finding suggested that stereochemically simpler *nor*-analogs of **1** might also inhibit glycoside hydrolysis. We now wish to report an extremely rapid and efficient synthesis of des(hydroxymethyl)-nojirimycin **2**. As expected, this achiral triol is in its own right a potent and highly selective almond  $\beta$ -glucosidase inhibitor. We have also prepared the mannose and galactose analogs **3** and **4**. These enantiomeric structures likewise inhibit glycosidases and shed new light on H-bonding and other enzyme-substrate interactions in the catalyzed hydrolysis of glycosides.



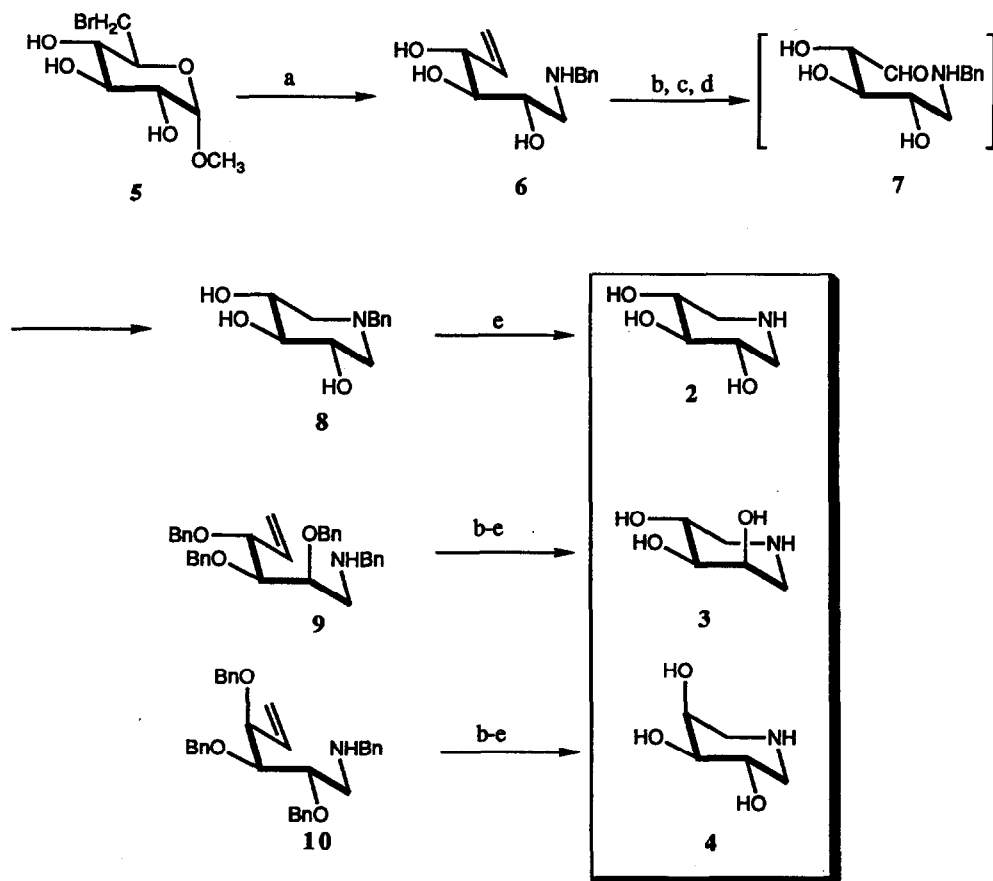
We planned a general synthesis of five-carbon iminoalditols like **2** from D-hexoses as outlined in the Scheme. The reductive ring opening of bromopyranose sugars, first developed by Vasella<sup>12</sup> and modified by us<sup>8</sup> to incorporate an *in situ* reductive amination of the incipient  $\omega$ -alkenylaldehyde, was successful in transforming methyl 6-bromo-6-deoxy- $\alpha$ -D-glucopyranoside<sup>13</sup> **5** into aminoalkenetriol **6** in a one-pot reaction (70% yield).<sup>14</sup> To our knowledge this represents the first example of direct, reductive opening of an unprotected glycoside. Moreover the reaction proceeded without complications arising from aldehyde epimerization or acetal/hemiacetal formation. To prevent oxidation of the benzylic amine in the next step,<sup>15</sup> ozonolysis of **6** was performed on the corresponding trifluoroacetate salt. When the first-formed ozonide was reductively cleaved using dimethylsulfide in the presence of sodium cyanoborohydride (2.5 equiv in CH<sub>3</sub>OH), trihydroxypiperidine **8** was obtained directly in 57% yield.<sup>16</sup> Hydrogenolysis of **8** quantitatively afforded the desired 1,5-dideoxy-1,5-imino-D-xylytol **2**.<sup>17</sup> In similar fashion D-mannose and D-galactose were converted via intermediates **9** and **10** to the enantiomerically related **3**<sup>18</sup> and **4**,<sup>19</sup> respectively.

At pH 5.0, alditol **2** inhibited sweet almond  $\beta$ -glucosidase (35% of control activity at 1 mM) but had no effect whatsoever on yeast  $\alpha$ -glucosidase, jackbean  $\alpha$ -mannosidase, coffee  $\alpha$ -galactosidase,  $\beta$ -galactosidase,  $\beta$ -glucuronidase, or  $\beta$ -hexosaminidase (all bovine). Kinetic measurements with **2** on almond  $\beta$ -glucosidase using *p*-nitrophenyl  $\beta$ -D-glucopyranoside as substrate ( $K_M = 2.5$  mM) resulted in Lineweaver-Burk plots indicating competitive inhibition.<sup>20</sup> Additionally, Dixon plots of  $1/V$  versus  $[I]$  gave a value of  $K_I$  for **2** ( $0.43 \pm 0.1$  mM) similar to that for **1** (0.37 mM at pH 5).<sup>11</sup> These data indicate that the hydroxymethylene sidechain of **1** is relatively unimportant for inhibitor binding.

Likewise, in exploratory screens against the same group of enzymes, mannose analog **3** competitively inhibited only jackbean  $\alpha$ -mannosidase. Its effect was comparable to that of 1-deoxymannojirimycin (40% of control activity at 1 mM). However galactose analog **4** had only marginal effects on  $\alpha$ - or  $\beta$ -galactosidase, suggesting that the  $-\text{CH}_2\text{OH}$  group of galactose may serve as a much more important recognition unit for catalyzed glycoside hydrolysis.

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## SCHEME



(a) excess Zn, 19:1 PrOH:H<sub>2</sub>O, NaBH<sub>3</sub>CN (2 equiv), PhCH<sub>2</sub>NH<sub>2</sub> (15 equiv), reflux, 2h;  
 (b) CH<sub>2</sub>Cl<sub>2</sub>, excess CF<sub>3</sub>CO<sub>2</sub>H, evaporation; (c) CH<sub>2</sub>Cl<sub>2</sub>, O<sub>3</sub> (1.1 equiv), -78°C; (d) (CH<sub>3</sub>)<sub>2</sub>S  
 (1.5 equiv), NaBH<sub>3</sub>CN (2.5 equiv), CH<sub>3</sub>OH, rt, 3h; (e) Pd-C, H<sub>2</sub>, CH<sub>3</sub>OH.

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14. For **6**:  $[\alpha]_D = -26^\circ$  ( $c=0.9$ ,  $\text{CHCl}_3$ );  $^1\text{H-NMR } \delta$  ( $\text{CDCl}_3$ ) 7.26-7.14 (m, 5 H), 5.82 (ddd, 1 H,  $J=17.2$ , 10.5, 5.7 Hz), 5.26 (dd, 1 H,  $J=17.2$ , 2.8 Hz), 5.12 (dd, 1 H,  $J=10.5$ , 2.8 Hz), 4.16 (m, 1 H), 3.74 (m, 1 H), 3.67 (d, 2 H,  $J=4.8$  Hz), 3.40 (dd, 1 H,  $J=4.0$ , 2.9 Hz), 2.79 (dd, 1 H,  $J=12.2$ , 4.1 Hz), 2.69 (dd, 1 H,  $J=12.2$ , 5.3 Hz); CMR ( $\text{CDCl}_3$ ) 139.2, 137.5, 128.5, 128.1, 127.2, 116.5, 75.4, 73.8, 70.4, 53.7, 51.9; IR (film) 3400, 2960, 1520, 1480, 1140, 1080, 1020  $\text{cm}^{-1}$ ; CIMS (isobutane)  $m/e$  238 ( $M+1$ , 100%)
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16. For **8**:  $[\alpha]_D = 0.0^\circ$ ;  $^1\text{H-NMR } \delta$  ( $\text{CD}_3\text{OD}$ ) 7.22-7.15 (m, 5 H), 3.46 (s, 2 H), 3.33 (m, 2 H), 2.98 (t, 1 H,  $J=8.9$  Hz), 2.83 (m, 2 H), 1.83 (m, 2 H); CMR ( $\text{CD}_3\text{OD}$ ) 138.7, 130.4, 129.3, 128.4, 80.6, 71.6, 63.1, 59.2; IR (KBr) 3330, 3270, 3050, 2935, 1450, 1365, 1145, 1075, 1020; CIMS  $m/e$  224 ( $M+1$ , 39%).
17. For **2**:  $[\alpha]_D = 0.0^\circ$ ;  $^1\text{H-NMR}$  ( $\text{D}_2\text{O}$ ) 3.56 (m, H-3, H-5), 3.33 (t, H-4,  $J=8.9$  Hz), 3.18 (dd, H-2<sub>eq</sub>, H-6<sub>eq</sub>,  $J=12.5$ , 4.8 Hz), 2.53 (dd, H-2<sub>ax</sub>, H-6<sub>ax</sub>,  $J=12.7$ , 12.2 Hz); CMR ( $\text{D}_2\text{O}$ ) 77.2, 69.8, 48.4; IR (KBr) 3400, 2920, 1645, 1455, 1075, 1045, 1025, 1010; CIMS  $m/e$  116 ( $M+1-\text{H}_2\text{O}$ , 100%).
18. For **3-HCl** salt (73% overall yield from the corresponding aminoalkene): mp 191-192°C;  $[\alpha]_D = -16^\circ$  ( $c=0.9$ ,  $\text{CH}_3\text{OH}$ );  $^1\text{H-NMR}$  ( $\text{D}_2\text{O}$ ) 4.26 (m, 1 H), 4.13 (ddd, 1 H,  $J=8.3$ , 7.7, 4.0 Hz), 3.81 (dd, 1 H,  $J=7.7$ , 2.7 Hz), 3.44 (dd, 1 H,  $J=12.8$ , 4.0 Hz), 3.33 (dd, 1 H,  $J=13.0$ , 5.9 Hz), 3.24 (dd, 1 H,  $J=13.0$ , 2.7 Hz), 2.99 (dd, 1 H,  $J=12.8$ , 8.3 Hz); IR (KBr) 3300 3080, 2930, 2910, 2860, 1615, 1480, 1450, 1410, 1370  $\text{cm}^{-1}$ ; CIMS  $m/e$  134 ( $M+1$ , 100%).
19. For **4-HCl** (82% overall yield from the corresponding aminoalkene): mp 191-192°C;  $[\alpha]_D = +16^\circ$  ( $c=0.5$ ,  $\text{CH}_3\text{OH}$ ); all other spectral data identical with **3-HCl**.
20. At very low concentrations ( $\leq 30 \mu\text{M}$ ), **2** behaved as a mild *activator* of  $\beta$ -glucosidase, judging from plots of absorbance versus  $[\text{I}]$  at several different substrate concentrations.

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